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Enhancement of Germination and Early Development of Sweet Corn Seeds Using Aerated Priming with KNO₃ and Reverse Osmosis Water

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Abstract. The -sh2, bt2, su1, and se- genes in sweet corn cause faster seed deterioration and a lower germination rate. A potential solution to this issue is seed priming. Priming agents and the solution uptake method are key factors that affect the achievement of seed priming. This study aims to compare the influence of solution uptake methods in sweet corn seed priming as well as investigate the effect of KNO3 on the germination and early development of sweet corn plants. The research was conducted in the Seed processing lab of the Department of Agricultural Production, Politeknik Negeri Jember. The seeds used in this study were sweet corn seeds var. Enno 1401. The study was designed with a completely randomized design (CRD). Seven treatments, namely non-primed, KNO3 spraying, KNO3 soaking, KNO3 aeration, RO water spraying, RO water soaking, and RO water aeration, were observed. Data were analyzed using one-way ANOVA, followed by means analysis using Fisher's LSD (least significant difference) test at P<0.05. The result showed that aeration priming performed better than spraying and soaking in terms of germination percentage, MGT, and GRI of sweet corn seeds. However, aeration priming with KNO3 gave lower results than RO water in terms of MGT and GRI. On the other hand, aeration priming with KNO3 aeration gave the highest result in dry weight.

Keywords: Halopriming, Hydropriming, Invigoration, Seed hydration

1. Introduction

Maize (*Zea mays* L.) is one of the most important food crops in the world used as a staple food, animal feed, oil, cereals, syrup, and other industrial products (Shah *et al.*, 2016). It is grown widely across the globe with an average production of 1.16 billion tons per year which equals to around 231.8 billion USD (FAOSTAT, 2023). Maize is known as a rich source of carbohydrates, fiber, minerals, and vitamins such as vitamins B1, B2, B3, B5, B6, B9, C, E, and K (Sharma *et al.*, 2022). It is remarkably adaptable and can grow in a variety of agroclimatic situations which leads to massive domestication and diverse variety development.

Among other maize varieties, sweet corn (*Zea mays* L. Var. *Saccharata*) is growing in popularity due to its sweeter taste and antioxidant activity. Sweet corn variety is basically maize that undergoes a mutation so that it collects twice as much sugar in the kernels as standard corn (Ngenoh *et al.*, 2015). These mutated genes prevent the alteration of sugar into starch in the

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endosperm which causes the accumulation of sugar in the kernels (Heryanto *et al.*, 2022). Four recessive genes—*sh2*, *bt2*, *su1*, and *se*—are the most frequent genes employed in breeding sweet corn (Santos *et al.*, 2014). However, due to the lower starch content caused by these genes, sweet corn seeds have lower vigor which leads to faster deterioration and lower germination rate (Lee, 2001).

One way to overcome this problem is by seed priming. Seed priming is the controlled hydration of seeds prior to sowing, during which the seeds start to germinate but are redried before reaching the stage of radicle/epicotyl extension (Bradford, 1986). Seed priming has proven to improve plant survivability in hostile conditions by providing coordinated, quick, and on-demand germination and establishment as well as tolerance to a variety of challenges of many commodities (Pedrini *et al.*, 2020), including maize. Some studies have reported the positive influence of seed priming on maize seedling performances using several priming agents, such as hydropriming PEG and GA₃ (Adhikari & Subedi, 2022), micronutrients (Nciizah *et al.*, 2020), salicylicacid (AL-Obaedi, 2022), as well as potassium nitrate (KNO₃) (Anosheh *et al.*, 2011). Since potassium nitrate aids in the formation of nitric oxide (NO), it is frequently used in seed priming (Hendricks & Taylorson, 1974), which breaks seed dormancy and promotes germination by interacting with reactive oxygen species (ROS), phytochrome signaling pathways, and ethylene production (Šírová *et al.*, 2011).

Besides priming agents, the solution uptake method is also a determining key that affects the achievement of seed priming. Thongtip *et al.* (2022) mentioned that seeds may experience oxygen deprivation when soaked in a solution for a certain time. Santika *et al.* (2022) discovered that aerated priming in tomato seed resulted in better Mean Germination Time (MGT) and Germination Rate Index (GRI) compared to soaking and spraying. Aerated priming is a seed priming method where the seeds are immersed in a priming solution and concurrently streamed with air coming from an air pump. This method ensures that the seeds are safe from the lack of oxygen during priming. However, there has not been any reports regarding aerated priming application in maize, especially in sweet corn seeds. Thus, this research aimed to compare the influence of solution uptake methods in sweet corn seed priming as well as to investigate the effect of KNO₃ on the germination and early development of sweet corn plant.

2. Methods

This study was conducted at the Seed Processing Laboratory, Politeknik Negeri Jember, from August to November 2022. The seeds of sweet corn tested in this study were var. Enno 1401. This variety was released in 2022 with a high potential yield and high sugar content at 11-13 °Brix.

The initial germination rate was at 79% (based on a preliminary study) which can still be improved further.

Two kinds of priming solutions were prepared: 0.75% KNO₃ and Reverse-Osmosis (RO) water. The priming methods studied were aeration, soaking, and spraying. A home-built aerator setup was used for the aeration method which was made by connecting plastic bottles, rubber tubes, and an electric air pump based on Santika et al. (2022). For the aeration procedure, 50 ml of the appropriate priming solution was placed in plastic bottles that were linked to an air pump, and the seeds were then allowed to aerate. To apply the soaking procedure, the seeds were placed in plastic bottles with 50 ml of the appropriate priming solutions inside of them, without any active aeration. The seeds were sprayed with the appropriate solutions and placed on germination paper using the spraying method. The seeds were primed for 24 hours in each treatment and exposed to 12 hours of light per day at 25 to 30 °C. Every treatment was carried out four times. After that, the seeds were allowed to air dry for three days in a public area. The dried seeds then are used in two studies: one on-field performance, and the other on germination. In the germination investigation, 100 seeds were used for each replication and germinated using the between-paper test method. The seedlings were then maintained at 25 to 30 °C for seven days (final count) under a 12-hour light cycle. For 42 days leading up to the field performance, the seeds were placed within polybags filled with a 2:1 mixture of topsoil and organic fertilizer.

The Final Germination Percentage (FGP), Mean Germination Time (MGT), and Germination Rate Index (GRI) were the observation parameters for the germination investigation. Only the final percentage of germination is reflected in FGP. A seed population's germination rate increases with its FGP value (Scott *et al.*, 1984). Germination in this definition based on ISTA Rules is the emergence of 2 mm of radicles. According to ISTA Rules, germination is defined as the appearance of 2 mm of radicles. The mean time required for seeds to germinate is known as MGT. A population of seeds germinated more quickly the lower the MGT (Orchard, 1977). The percentage of germination on each day of the germination phase is shown in the GRI. Higher and quicker germination is indicated by higher GRI values (Esechie, 1994). For the germination investigation, the plant height (cm), stem diameter (cm), and dry mass (g) were the observation parameters.

The design of the experiment was completely randomized (CRD). One-way ANOVA was performed for analyzing all the data and means analysis using Fisher's LSD (least significant difference) test at P<0.05 followed after.

3. Results and Discussion

In this study, two types of solution (RO water and 0.75% KNO₃) were used as seed priming agents for sweet corn. Based on Table 1, the RO water solution is better than KNO3 in terms of MGT and GRI. Several studies on various plant species confirm that hydro priming can increase the germination rate index, as well as shorter MGT (Moreno et al., 2018; Alias et al., 2018; Meena et al., 2001; Ramzan et al., 2010; Ghassemi-Golezani et al., 2010). However, the two solutions show a similar pattern where the aeration method produces better results than soaking and spraying in terms of MGT and GRI (Table 1). In tomato seed priming, aeration tends to increase germination percentage, MGT, and GRI both in KNO3 and Water solution (Santika et al., 2022). Aerated solution seeds tended to absorb more water, suggesting that oxygen promotes water uptake (Yeoung et al., 1995). One disadvantage of using the spraying method in conjunction with hydropriming is that it may occasionally lead to uneven hydration, which in turn produces un-uniform germination. Certain plant species with thin coats such as sweet corn could not benefit from it, where quick hydration can cause seed damage generated by nutrient leakage from the seed (Adnan et al., 2020). The solution's composition and osmotic potential, as well as the time, temperature, and level of aeration, all influence the seed priming response (Nascimento, 2003). During seed priming, oxygen is necessary (Heydecker et al., 1975; Bujalski et al., 1989). While some studies (Bradford et al., 1988; Nerson & Govers, 1986), recommend aeration during melon seed priming, others note negative consequences. Aerating the solution reduces the amount of time needed to prime lettuce seeds (Guedes & Cantliffe, 1981).

Table 1. Average of Germination Percentage, MGT, and GRI of primed sweet corn seeds

Treatments	Germination (%)	MGT (day)	GRI
Combination			
Non-primed	$79.50 \pm 1.04 c$	2.62 <u>+</u> 0.08 c	38.52 ±1.48 a
KNO ₃ Spraying	$80.25 \pm 1.18 \mathrm{c}$	$2.73 \pm 0.09 c$	38.94 <u>+</u> 1.35 a
KNO ₃ Soaking	$76.00 \pm 1.22 \mathrm{b}$	$2.55 \pm 0.05 c$	35.26 ±1.39 a
KNO ₃ Aeration	74.75 <u>+</u> 1.11 b	2.24 <u>+</u> 0.06b	37.04 ±1.42 a
RO Water Spraying	$61.75 \pm 1.03 a$	2.59 <u>+</u> 0.09 c	27.53 ±1.06 a
RO Water Soaking	$79.75 \pm 1.31 c$	$2.36 \pm 0.07 \mathrm{bc}$	39.18 <u>+</u> 1.40 a
RO Water Aeration	84.00 <u>+</u> 1.08 d	1.86 <u>+</u> 0.08 a	59.75 <u>+</u> 1.58 b
LSD	3.37	0.22	4.09
Solution			
RO Water	75.17 ±1.73 a	2.27 ±0.08 a	45.48 <u>+</u> 3.13 b
0.75% KNO3	77.00 <u>+</u> 0.94 a	2.51 <u>+</u> 0.07 b	37.08 <u>+</u> 0.85 a
LSD	4.08	0.22	6.73
Solution Uptake Method			
Soaking	77.75 ±2.23 ab	2.45 ±0.07 b	48.39 <u>+</u> 1.17 a
Spraying	71.00 ±2.41 a	2.66 ±0.06b	38.24 <u>+</u> 0.84 a
Aeration	79.38 <u>+</u> 2.27 b	$2.05 \pm 0.08 a$	48.39 <u>+</u> 4.40 b
LSD	6.78	0.22	1.97

Remarks:

Means within each cell followed by different letters are significantly different according to Fisher's Least Significant Difference test at $P \le 0.05$.

Table 2. Average ofplant height, stem diameter, and dry mass of primed sweet corn seeds

Treatments	plant height	stem diameter	dry mass
Treatments	at 28 DAP (cm)	at 28 DAP (mm)	(g)
Combination			
Non-primed	43.73 <u>+</u> 4.77 a	18.83 <u>+</u> 0.92 b	176.1 <u>+</u> 5.0 ab
KNO ₃ Spraying	42.71 <u>+</u> 4.59 a	17.33 <u>+</u> 0.83 ab	181.4 <u>+</u> 4.7 b
KNO ₃ Soaking	49.87 <u>+</u> 3.05 a	19.61 <u>+</u> 0.74 b	195.4 ±4.4 bc
KNO ₃ Aeration	44.94 <u>+</u> 4.20 a	19.03 <u>+</u> 0.78 b	204.0 <u>+</u> 4.5 c
RO Water Spraying	39.43 <u>+</u> 3.23 a	16.03 <u>+</u> 0.84 a	184.9 <u>+</u> 5.4 b
RO Water Soaking	47.09 <u>+</u> 1.89 a	20.08 <u>+</u> 0.76 b	169.3 <u>+</u> 6.4 ab
RO Water Aeration	39.43 <u>+</u> 4.42 a	19.04 <u>+</u> 0.68 b	165.4 <u>+</u> 5.5 a
LSD	11.35	2.34	15.2
Solution			
RO Water	43.50 ±1.99 a	18.38 <u>+</u> 0.65 a	173.2 <u>+</u> 4.0 a
KNO3	45.84 <u>+</u> 2.28 a	18.65 <u>+</u> 0.50 a	193.6 <u>+</u> 3.7 b
LSD	6.27	1.71	11.2
Solution Uptake Method			
Soaking	48.48 ±1.74 b	19.85 <u>+</u> 0.50 b	182.4 <u>+</u> 6.1 a
Spraying	44.46 <u>+</u> 2.83 a	16.68 <u>+</u> 0.60 a	183.2 <u>+</u> 3.4 a
Aeration	41.07 <u>+</u> 2.67 ab	19.03 <u>+</u> 0.48 b	184.7 <u>+</u> 6.0 a
LSD	7.24	1.55	15.61

Remarks:

Means within each cell followed by different letters are significantly different according to Fisher's Least Significant Difference test at $P \le 0.05$.

There were no significant differences in corn plant height on all treatments (Table 2). In terms of stem diameter (28 DAP), both RO water and KNO3 showed higher results than aeration when applied by aeration or soaking than the spraying method. The smallest stem diameter (28 DAP) was obtained by RO water spraying (16.03 ± 0.84 mm) which was not significantly different from KNO₃ spraying (17.33 \pm 0.83 mm). The seed damage caused by uneven hydration and nutrient leakage caused by the spraying method during germination is thought to have an impact later on the disruption of sweet corn stem growth. In dry mass, KNO3 aeration recorded the highest number (204,0 \pm 4,5 g) while RO water aeration showed the lowest number (165,4 \pm 5,5 g). Although RO water aeration shows the best numbers in germination percentage, MGT, and GRI (Table 1), it is inversely proportional to its dry mass. This might due to fact that KNO₃ contains potassium and nitrogen which act as nutrients in boosting the seedling metabolism resulting in better dry weight. In some cases, aeration during hydropriming can trigger the emergence of pathogenic fungi which can disturb the growth of plants (Nascimento, 2003). In other reports, the seeds primed with KNO3 could demonstrate superior germination rate and seedling dry weight, as well as the most standard attributes for all parameters as compared to the control group (Ghobadi et al., 2012; Ahmadvand et al., 2012; Mohammadi & Amiri, 2010). For several kinds of crops, seed priming with nitrate salts can regulate essential growth characteristics even in unfavorable environmental circumstances (Thakur et al., 2019). As growth regulators to plants, nitrate salts boost the amount of photosynthetic assimilates that are translocated to areas that contribute to production (Srivastava *et al.*, 2017).

4. Conclusion

It can be concluded that aeration priming performed better than spraying and soaking in terms of germination percentage, MGT, and GRI of sweet corn seeds. However, dissimilarly with other findings, aeration priming with KNO₃ gave lower results than RO water in terms of MGT, GRI. On the other hand, aeration priming with KNO₃ aeration gave the highest result in dry weight. Further researches regarding different type of solutes used in aeration priming, as well as the biochemical properties and the yield of primed sweet corn in the field need to be carried out.

References

- Adhikari, S., & Subedi, R. (2022). Effect Of Seed Priming Agents (GA3, PEG, Hydropriming) In The Early Development Of Maize. *Russian Journal of Agricultural and Socio-Economic Sciences*, 9(129), 113–120. https://rjoas.com/issue-2022-09/article_11.pdf
- Adnan, M., Abd-ur-Rahman, H., Asif, M., Hussain, M., Bilal, M. B., Adnan, M., ..., & Khalid, M. (2020). Seed priming; An effective way to improve plant growth. EC Agriculture, 6(6), 01–05. https://www.researchgate.net/profile/Fazal-Ur-Rehman-2/publication/344887216_Seed_Priming_An_Effective_Way_to_Improve_Plant_Growth/links/61973c1ad7d1af224b070549/Seed-Priming-An-Effective-Way-to-Improve-Plant-Growth.pdf
- Ahmadvand, G., Soleymani, F., Saadatian, B., & Pouya, M. (2012). Effects of Seed Priming on Seed Germination and Seedling Emergence of Cotton Under Salinity Stress. *World Applied Sciences Journal*, 20(11), 1453–1458. https://www.researchgate.net/profile/Bijan-Saadatian/publication/288457838_Effects_of_seed_priming_on_seed_germination_and_seedling_emergence_of_cotton_under_salinity_stress/links/5ce5a7ce458515712ebb7d0a/Effects-of-seed-priming-on-seed-germination-and-seedling-emergence-of-cotton-under-salinity-stress.pdf
- AL-Obaedi, A. I. (2022). Evaluation of the Effect of Different Priming Treatments on the Seed Germination of Maize (Zea mays. L) Based on In Vitro Conditions. *Samarra Journal of Pure and Applied Science*, 4(1), 71–80. https://doi.org/10.54153/sjpas.2022.v4i1.345
- Alias, N. S. B., Billa, L., Muhammad, A., & Singh, A. (2018). Priming and temperature effects on germination and early seedling growth of some Brassica spp. *Acta Horticulturae*, 1225, 407–414. https://doi.org/10.17660/ActaHortic.2018.1225.57
- Anosheh, H. P., Sadeghi, H., & Emam, Y. (2011). Chemical priming with urea and KNO3 enhances maize hybrids (Zea mays L.) seed viability under abiotic stress. *Journal of Crop Science and Biotechnology*, 14(4), 289–295. https://doi.org/10.1007/s12892-011-0039-x
- Bradford, K. J. (1986). Manipulation of Seed Water Relations Via Osmotic Priming to Improve Germination Under Stress Conditions. *HortScience*, 21(5), 1105–1112. https://doi.org/10.21273/HORTSCI.21.5.1105
- Bradford, K. J., May, D. M., Hoyle, B. J., Skibinski, Z. S., Scott, S. J., & Tyler, K. B. (1988). Seed and Soil Treatments to Improve Emergence of Muskmelon from Cold or Crusted Soils. *Crop Science*, 28(6), 1001–1005. https://doi.org/10.2135/cropsci1988.0011183X002800060028x
- Bujalski, W., Nienow, A. W., & Gray, D. (1989). Establishing The Large Scale Osmotic Priming of Onion Seeds by Using Enriched Air. *Annals of Applied Biology*, 115(1), 171–176. https://doi.org/10.1111/j.1744-7348.1989.tb06824.x
- Esechie, H. A. (1994). Interaction of Salinity and Temperature on the Germination of Sorghum. Journal of Agronomy and Crop Science, 172(3), 194–199. https://doi.org/10.1111/j.1439-

037X.1994.tb00166.x

- FAOSTAT. (2023). Crops and livestock products. https://www.fao.org/faostat/en/#data/QCL
- Ghassemi-Golezani, K., Chadordooz-Jeddi, A., Nasrollahzadeh, S., & Moghaddam, M. (2010). Effects of Hydro-Priming Duration on Seedling Vigour and Grain Yield of Pinto Bean (Phaseolus vulgaris L.) Cultivars. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 38(1), 109–113. https://doi.org/https://doi.org/10.15835/nbha3813475
- Ghobadi, M., Abnavi, M. S., Jalali-Honarmand, S. Mohammadi, G. R., & Ghobadi, E. (2012). Effects of seed priming with some plant growth regulators (Cytokinin and salicylic acid) on germination parameters in wheat (Triticum aestivum L.). *Journal of Agricultural Technology*, 8(7), 2157–2167. http://www.aatsea.org/images/conference_publications/pdf/v8_n7_12_December/2_IJAT_2012_8(7) Shafiei Abnavi M Agronomy.pdf
- Guedes, A. C., & Cantliffe, D. J. (1981). Germination of Lettuce Seeds at High Temperature After Seed Priming. *Journal of the American Society for Horticultural Science*, 105(6), 777–778. https://www.cabidigitallibrary.org/doi/full/10.5555/19810391327
- Hendricks, S. B., & Taylorson, R. B. (1974). Promotion of Seed Germination by Nitrate, Nitrite, Hydroxylamine, and Ammonium Salts. *Plant Physiology*, 54(3), 304–309. https://doi.org/10.1104/pp.54.3.304
- Heryanto, F. S. S., Wirnas, D., & Ritonga, A. W. (2022). Diversity of twenty-three sweet corn (Zea mays L. saccharata) varieties in Indonesia. *Biodiversitas Journal of Biological Diversity*, 23(11), 6075–6081. https://doi.org/10.13057/biodiv/d231164
- Heydecker, W., Higgis, J., & Turner, Y. J. (1975). Invigoration of seeds. *Seed Science & Technology*, 3, 881–888. https://cabidigitallibrary.org/doi/full/10.5555/19760342303
- Lee, M.-H. (2001). Seed Deterioration Response of Different Genes of Sweet Corn during Longterm Storage. *Korean Journal of Crop Science*, 46(4), 317–320. https://koreascience.kr/article/JAKO200111922228558.page
- Meena, R. P., Tripathi, S. C., Chander, S., Chhokar, R. S., & Sharma, R. K. (2001). Seed priming in moisture-stress conditions to improve growth and yield of wheat (Triticum aestivum). *Indian Journal of Agronomy*, 60(1), 99–103. https://doi.org/10.59797/ija.v60i1.4421
- Mohammadi, G. R., & Amiri, F. (2010). The Effect of Priming on Seed Performance of Canola (Brassica napus L.) under Drought Stress. *American-Eurasian Journal of Agricultural and Environmental Science*, 9(2), 202–207. https://www.cabidigitallibrary.org/doi/full/10.5555/20113179677
- Moreno, C., Seal, C. E., & Papenbrock, J. (2018). Seed priming improves germination in saline conditions for Chenopodium quinoa and Amaranthus caudatus. *Journal of Agronomy and Crop Science*, 204(1), 40–48. https://doi.org/10.1111/jac.12242
- Nascimento, W. M. (2003). Muskmelon seed germination and seedling development in response to seed priming. *Scientia Agricola*, 60(1), 71–75. https://doi.org/10.1590/S0103-90162003000100011
- Nciizah, A. D., Rapetsoa, M. C., Wakindiki, I. I., & Zerizghy, M. G. (2020). Micronutrient seed priming improves maize (*Zea mays*) early seedling growth in a micronutrient deficient soil. *Heliyon*, 6(8), e04766. https://doi.org/10.1016/j.heliyon.2020.e04766
- Nerson, H., & Govers, A. (1986). Salt Priming of Muskmelon Seeds for Low-temperature Germination. *Scientia Horticulturae*, 28(1–2), 85–91. https://doi.org/10.1016/0304-4238(86)90127-5
- Ngenoh, E., Mutai, B. K., Chelang'a, P. K., & Koech, W. (2015). Evaluation of Technical Efficiency of Sweet Corn Production among Smallholder Farmers in Njoro district, Kenya. *Journal of Economics and Sustainable Development*, 6(17), 183–193. https://core.ac.uk/download/pdf/234647227.pdf
- Orchard, T. J. (1977). Estimating the parameters of plant seedling emergence. *Seed Science and Technology*, 5(1), 61–69. https://eurekamag.com/research/000/368/000368846.php

- Pedrini, S., Balestrazzi, A., Madsen, M. D., Bhalsing, K., Hardegree, S. P., Dixon, K. W., & Kildisheva, O. A. (2020). Seed enhancement: getting seeds restoration-ready. *Restoration Ecology*, 28(S3), S266–S275. https://doi.org/10.1111/rec.13184
- Ramzan, A., Hafiz, I. A., & Abbasi, N. A. (2010). Effect Of Priming with Potassium Nitrate and Dehusking on Seed Germination of Gladiolus (Gladiolus Alatus). *Pakistan Journal of Botany*, 42(1), 247–258. http://pakbs.org/pjbot/PDFs/42(1)/PJB42(1)247.pdf
- Santika, P., Muhklisin, I., & Makama, S. D. (2022). Effect of Aeration and KNO3 in Seed Priming on The Germination of Tomato (Solanum lycopersicum) Seeds. *Agroteknika*, 5(2), 151–160. https://doi.org/10.55043/agroteknika.v5i2.153
- Santos, P. H. A. D., Pereira, M. G., Trindade, R. dos S., Cunha, K. S. da, Entringer, G. C., & Vettorazzi, J. C. F. (2014). Agronomic performance of super-sweet corn genotypes in the north of Rio de Janeiro. *Crop Breeding and Applied Biotechnology*, 14(1), 8–14. https://doi.org/10.1590/S1984-70332014000100002
- Scott, S. J., Jones, R. A., & Williams, W. A. (1984). Review of Data Analysis Methods for Seed Germination. *Crop Science*, 24(6), 1192–1199. https://doi.org/10.2135/cropsci1984.0011183X002400060043x
- Shah, T., Prasad, K., & Kumar, P. (2016). Maize-A potential source of human nutrition and health:

 A review. Cogent Food & Agriculture, 2(1). https://doi.org/10.1080/23311932.2016.1166995
- Sharma, Y., Wadhawan, N., & Lakhawat, S. (2022). Analysis of nutritional composition of popular maize varieties. *The Pharma Innovation Journal*, 11(10), 238–241. https://www.thepharmajournal.com/archives/2022/vol11issue10S/PartD/S-11-9-318-387.pdf
- Šírová, J., Sedlářová, M., Piterková, J., Luhová, L., & Petřivalský, M. (2011). The role of nitric oxide in the germination of plant seeds and pollen. *Plant Science*, 181(5), 560–572. https://doi.org/10.1016/j.plantsci.2011.03.014
- Srivastava, A. K., Siddique, A., Sharma, M. K., & Bose, B. (2017). Seed Priming with Salts of Nitrate Enhances Nitrogen use Efficiency in Rice. Vegetos- An International Journal of Plant Research, 30(4), 99-104. https://doi.org/10.5958/2229-4473.2017.00199.9
- Thakur, M., Sharma, P., & Anand, A. (2019). Seed Priming-Induced Early Vigor in Crops: An Alternate Strategy for Abiotic Stress Tolerance. In *Priming and Pretreatment of Seeds and Seedlings* (pp. 163–180). Springer Singapore. https://doi.org/10.1007/978-981-13-8625-1_8
- Thongtip, A., Kaewsorn, A., & Chatbanyong, R. (2022). Effects of KNO3 Concentration and Aeration during Seed Priming on Seed Germination and Vigor of Papaya cv. Khaek Dam Kaset. *Rajamangala University of Technology Srivijaya Research Journal*, *14*(1), 1–15. https://li01.tci-thaijo.org/index.php/rmutsvrj/article/view/242427
- Yeoung, Y. R., Wilson, D. J., & Murray, G. A. (1995). Oxygen regulates imbibition of muskmelon seeds. *Seed Science & Technology*, 23, 843–850. https://www.cabidigitallibrary.org/doi/full/10.5555/19960304093

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